



Article

Local Genomic Instability of the *SpTransformer* Gene Family in the Purple Sea Urchin Inferred from BAC Insert Deletions

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Abstract: The *SpTransformer* (*SpTrf*) gene family in the purple sea urchin, *Strongylocentrotus purpuratus*, encodes immune response proteins. The genes are clustered, surrounded by short tandem repeats, and some are present in genomic segmental duplications. The genes share regions of sequence and include repeats in the coding exon. This complex structure is consistent with putative local genomic instability. Instability of the *SpTrf* gene cluster was tested by 10 days of growth of *Escherichia coli* harboring bacterial artificial chromosome (BAC) clones of sea urchin genomic DNA with inserts containing *SpTrf* genes. After the growth period, the BAC DNA inserts were analyzed for size and *SpTrf* gene content. Clones with multiple *SpTrf* genes showed a variety of deletions, including loss of one, most, or all genes from the cluster. Alternatively, a BAC insert with a single *SpTrf* gene was stable. BAC insert instability is consistent with variations in the gene family composition among sea urchins, the types of *SpTrf* genes in the family, and a reduction in the gene copy number in single coelomocytes. Based on the sequence variability among *SpTrf* genes within and among sea urchins, local genomic instability of the family may be important for driving sequence diversity in this gene family that would be of benefit to sea urchins in their arms race with marine microbes.

Keywords: *Strongylocentrotus purpuratus*; short tandem repeats; tandem duplications; long-read assembly; large DNA deletions



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1. Introduction

There are many examples of gene copy number expansion into immune response families that function in innate immune systems among organisms. Many examples include cell surface, cytoplasmic, and secreted pathogen recognition receptors that bind microbes and/or their molecules and activate the innate immune response in the presence of possible pathogens or opportunists (e.g., [1–3]). The resulting gene families are under selection pressure for genes to be retained or to be deleted from the genome and perhaps to diversify based on pathogens and/or environmental variations. The outcome for the increased size and structure of the gene families results in benefits to the host for a broad recognition of the associated microbial populations and survival in its environment. However, the underlying mechanisms for the initial expansion of a single gene copy into a family is not understood. We have proposed previously that genomic instability is part of the complexity that results in gene expansion [4]. Genomic instability (also referred to as genomic fragility) has been the subject of many studies, a significant number debating its origin. Instability is associated with a high density of genomic sequence repeats [5], which are considered to be

rapidly evolving compared to the rest of the genome [6]. These repeats are thought to be generated through processes such as conversion, recombination, slipping misalignments, and single-strand annealing [6,7]. Long regions of repeats can result in secondary structures, such as Z-DNA, bulge loops, hairpin loops, four loop junctions, and G-quadruplexes, that may impair cellular processes (reviewed in [6]). The impairment of cellular processes, including DNA replication and repair, can lead to segmental duplications [6,8–11], elevated recombination rates [12,13], duplications of tRNA genes [14,15], non-homogeneity of gene distribution [16], and expanded regulatory regions [17,18].

The *Transformer* (*Trf*) gene family (previously termed the 185/333 genes) encodes immune response proteins and has been identified in several species of euechinoids [19–22]. The *SpTrf* genes are present as a family in the purple sea urchin and are upregulated swiftly by phagocytes in adults responding to several different pathogen-associated molecular patterns [23–25] and marine bacteria [26–30], and by filipodial cells in the blastocoel of larvae [31]. Several aspects of the structure of the genes and the *SpTrf* family in the purple sea urchin, *Strongylocentrotus purpuratus*, are consistent with genomic instability. The genes are small, with two exons, of which the second shows significant sequence diversity based on a variety of attributes [19]. It is composed of blocks of similar but non-identical sequences termed elements that are present in mosaic structures that are different among different genes (Figure 1A). Both tandem and interspersed repeats are also present in the second exon. The hypothetical evolutionary history of the tandem repeats suggests local duplications, ectopic duplications and insertions, deletions, and repeat recombination that resulted in two to four imperfect tandem repeats in the extant family [32,33]. In addition to the theoretical recombination among repeats, evidence of possible recombination among genes has also been reported [33]. Although there is significant sequence variability among the genes, they are 88% similar [19]. Consequently, the genes themselves may also be viewed as repeats. The repeats associated with the gene family, along with the sequence similarities among the genes themselves, are consistent with a region of local genomic instability in which evolutionary processes such as recombination may have key functions in sequence diversification and the evolution of both the genes and the family.

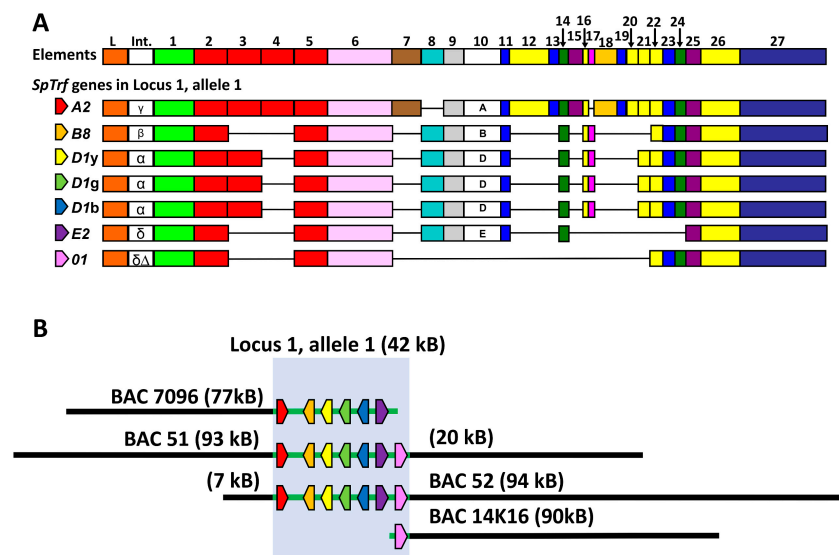


Figure 1. Graphical representation of BAC inserts that contain *SpTrf* gene locus 1, allele 1. (A) A graphical alignment of selected *SpTrf* genes showing the element pattern of each gene according to the repeat-based alignment [19]. Genes are composed of two exons; the first encodes the leader (L), and the second encodes the mature protein. All genes have a single, short intron (int) with several identifiable sequences that are indicated by the letters [19]. Exon 2 is a mosaic of elements (all of which are shown with numbers at the top) and is variable among the genes. Element 10 is labeled with different letters that indicate different sequences and define the element pattern. The horizontal

black lines indicate missing elements. This figure is modified from Figure 6B in [34]. (B) The overlaps among the BAC inserts containing allele 1 from locus 1 are highlighted with the blue box. The black lines to the left and right of the blue box indicate the size of the flanking regions of the inserts that are not part of the gene cluster. This figure is modified from Figure 4A in [35].

The characteristics of the *SpTrf* gene family structure are also consistent with genomic instability. The sequenced family, consisting of 17 alleles, is arranged in the sequenced genome (version 5.0) in two loci [36,37]. The genes are clustered and tightly linked, with intergenic distances ranging from 3.0 kb to 12 kb [32,35,38]. Locus 1 is composed of seven alleles on one chromosome and a mismatch of six alleles on the other (Figure 1B), whereas locus 2 has two genes, each with associated alleles [32]. Short tandem repeats (STRs) composed of two nucleotides, GA, surround each gene, and three regions of GA repeats of 3 kb to 4 kb are present in locus 2 in locations that appear to correspond with the locations of genes in locus 1 [32,35]. A different STR with the repeated sequence of GAT is present in multiple locations within both loci. In addition to the variety of repeats within and surrounding the genes, segmental duplications of 4.3 kb are also present that include duplicated genes, some so recent that their sequences are nearly identical [38], whereas other duplications have quite different sequences [32]. The genome assembly and the variety of BAC inserts that have been sequenced illustrate that the *SpTrf* family loci are riddled with a wide variety of repeats [32,35,37,38].

Genomic instability has been suggested previously, based on the descriptions of the genes and the *SpTrf* gene family structure described above [4]. Quantitative PCR was used to estimate the gene copy numbers in genomes from individual sea urchins, which indicated about 40 to 60 genes per genome [25]. However, the current genome assembly (version 5.0) shows only nine genes in two loci, many of which are annotated incorrectly ([Echino-base.org](https://echino-base.org), accessed on 26 October 2023). This underrepresentation has been attributed in part to a computational assembly problem that recognizes the multiple *SpTrf* genes as repeats or alleles and collapses them into single consensus genes, which appear as mixed sequences of two or more alleles or genes [35,38]. Furthermore, the *SpTrf-01* gene identified in a BAC insert sequence and a member of allele 1 in locus 1 [35] is not present in the genome. In an effort to overcome these assembly problems and present a corrected gene family structure, the BAC library that was sequenced for genome assembly (13X coverage of the genome; [39]) was screened for clones harboring *SpTrf* sequences [35]. It was notable that although 75 clones screened positive for *SpTrf* sequences, only 27 BAC clones supported PCR amplification of *SpTrf* sequences after *E. coli* was grown for BAC DNA isolation and analysis. Although this difference might have been the result of a failure of the PCR to amplify many genes for a variety of possible reasons, we propose that an alternative explanation is the instability of the BAC inserts based on the repeats and tight gene clustering that resulted in DNA deletions. This notion is in agreement with the underrepresentation of the *SpTrf* gene copy number in the sequenced genome that was assembled from overlapping BAC insert end sequences [37,40,41]. This hypothesis was tested with repeated inoculation and growth of *E. coli* harboring BAC clones for 10 days. BAC DNA with multiple *SpTrf* genes that was isolated from single colonies after the 10-day growth period had a variety of deletions. Alternatively, the BAC insert with only a single *SpTrf* gene was stable. Based on these results, we propose that local genomic instability is active in the *SpTrf* gene family in *S. purpuratus* and may be under cellular control to block deletion of the entire gene family. We propose that it is an underlying mechanism for the variability among the genes in the family, which includes sequence diversification that is a benefit for sea urchins in the arms race with marine microbial pathogens.

2. Methods

2.1. BAC Plasmids, DNA Isolation, and Sequencing

BAC clones were chosen based on their *SpTrf* gene copy numbers (Table 1) [35], which ranged in gene content from a single gene to the full cluster of seven genes in locus 1,

allele 1 (Figure 1A). *E. coli* harboring pBACe3.6 plasmids with sea urchin genomic (g)DNA inserts were spread on LB plates with 12.5 µg/mL chloramphenicol (LB/chlor) and grown overnight at 37 °C. Single colonies were inoculated into 5 mL of LB/chlor media and grown overnight at 37 °C with rotation. Bacteria in 1 µL of media were re-inoculated in 5 mL fresh media and grown overnight, which was repeated for 10 days. Bacteria in the final broth culture were spread on LB/chlor plates, single colonies (34–40) were selected per BAC and expanded in LB/chlor media, and BAC plasmid DNA was isolated by the alkaline lysis method as described in [19]. All control BACs (BAC-con) were isolated from *E. coli* harboring the pBACe3.6 plasmids with sea urchin gDNA inserts after a single day of growth as described above. BAC DNA was digested with *NotI* to release the insert from the plasmid and in combination with *XhoI* or *SacII*, and the inserts were evaluated by pulsed field gel electrophoresis (PFGE; CHEF-DR II Chiller System, Bio-Rad item #1703727) with 1% pulsed field certified agarose (Bio-Rad Laboratories, Hercules, CA, USA) in 0.3X concentration Tris borate EDTA (TBE) gel running buffer (27 mM Tris (pH 7.4), 27 mM boric acid, 0.6 mM EDTA). The PFGE parameters were 6 V/cm, with a switch time of 1–15 s over 16 h [35]. After separation, the gels were soaked in ethidium bromide solution and imaged with a UV imaging system (Gel Logic 1500 Imaging System, Kodak, Rochester, NY, USA). The locations of possible deletions were predicted by virtual restriction enzyme digests (<https://nc3.neb.com/NEBcutter>; accessed on 18 May 2018; New England Biolabs, Ipswich, MA, USA) based on the full-length insert sequence of each BAC, based on a previous report [42] or as reported here.

Table 1. BACs with multiple *SpTrf* genes show changes in insert sizes and gene copy numbers after a 10-day growth period.

BAC Clone Name ¹	GenBank Accession Number	BAC Insert Screens		SpTrf Gene Copies in BAC Inserts		
		Colonies Screened for BAC Insert Size	BACs with Deletions	Colonies Screened	Gene Copies Expected	Gene Copies in 8 BACs with Short Inserts
BAC-7096	BK007096	40	4 (10%)	8	3	3, 2, 2, 3, 3, 3, 3, 3
BAC-51	KU668451	34	1 (3%)	8	4	4, 4, 4, 3, 4, 4, 4, 4
BAC-52	KU668452	40	3 (7.5%)	8	4	4, 0, 4, 0, 4, 1, 4, 2
BAC-67	PP082967	40	0 (0%)	8	1	1, 1, 1, 1, 1, 1, 1, 1

¹ Abbreviated BAC accession numbers are used as clone names.

The *SpTrf* genes in the BAC inserts were identified by PCR amplicon sizes using primers that amplified all *SpTrf* genes based on annealing sites in the untranslated regions (5'UTR-forward: TAGCATCGGAGAGACCT; 3'UTR-reverse: AAATTCTACACCTCGGC-GAC), as described previously [25]. Amplicons were separated by gel electrophoresis and visualized with the Kodak UV imaging system.

2.2. BAC Insert Assembly from Sequencing Reads, Alignments, and Dot Plots

BAC DNA ($n = 3$) from single colonies was isolated after 10 days of re-inoculations for clones that showed smaller inserts compared to the full-length original BACs. In addition, BAC-con DNA ($n = 2$) from single colonies was isolated after a single day of growth and showed no insert size differences compared to the full-length original BAC inserts. BACs were grown in LB/chlor media overnight at 37 °C, and the plasmid DNA was isolated by a NucleoBond BAC100 high molecular weight DNA kit (Machery-Nagel Inc., Allentown, PA, USA) and evaluated for *E. coli* genomic DNA contamination by PCR (primers: Ecoli-1F, CGAAGCGACTGGAGCATGTG; Ecoli-1R, ACGCCACATTCGC-CAATTC) compared to amplicons of the plasmid (pBACe3.6F, AGCCGTGTAACCGAG-CATAGC; pBACe3.6R, GGAACATGACGGTATCTGCGAG). The reaction used PrimeSTAR GXL DNA polymerase (Takara Bio USA, San Jose, CA, USA), and the PCR program was an initial DNA melt at 98 °C for 30 s, 30 cycles of 98 °C for 10 s, 60 °C for 15 s, and 68 °C for 1 min, with a 68 °C extension and 4 °C hold. The amplicons for both pairs of

primers were the same size, and inserts from BAC DNA with low levels of contaminating genomic DNA from *E. coli* were sequenced at the University Maryland Institute for Genome Sciences (<https://marylandgenomics.org/>) using long-read technology (Pacific Biosciences, Menlo Park, CA, USA). BAC insert sequences were assembled using Canu version 2.2 (HiCanu; <https://github.com/marbl/canu/releases>, accessed on 27 September 2022) [43–45] using the following parameters: genome size = 0.03 M, corErrorRate = 0.045, batOptions = “–eg 0.0 –sb 0.001 –dg 3 –dr 0 –ca 2000 –cp 200”, and mhapipe = false.

Contigs covering each of the BAC inserts that were returned from the HiCanu assembly were aligned by hand in Molecular Evolutionary Genetics Analysis X (MEGAX) against the relevant original BAC insert sequence [35], and a consensus sequence was generated using EMBOSS Cons (https://www.ebi.ac.uk/Tools/msa/emboss_cons/ accessed on 27 September to 15 November 2023) from contigs returned from HiCanu assembly. The consensus sequence was used for further analysis. Sequence assemblies of BAC inserts were verified by BLAST searches of the sequencing reads against the original BAC sequences (GenBank accession numbers KU668451 and KU668452) that have been reported previously [35,38]. Insert sequences for BAC-51-15 and BAC-52-2b are available from GenBank (accession numbers PP082968 and PP082969, respectively). Sequence reads for BAC-42 and BAC-44 are available as raw sequence reads from GenBank (BioSample accession numbers SAMN39322606 and SAMN39322605, respectively).

Dot plots were generated using the YASS genomic similarity search tool (<https://bioinfo.univ-lille.fr/yass/index.php>, accessed on 3 October 2023) [46] to visualize the deletions in each BAC with a short insert against the original BAC insert sequence. The e-value threshold was set to e^{-30} , with the rest of the parameters left at standard settings (scoring matrix: +5, −4, −3, −4; gap costs: −16, −4; X-drop threshold: 30).

Raw PacBio reads for the BAC inserts were mapped against the relevant original BAC insert sequence [35] using minimap2 2.1 with preset parameters [47,48]. The output file was converted from a .sam file to a .bam file, sorted, and indexed using samtools 1.6 [49] before visualization in The Integrative Genomics Viewer (version 2.15.2) [50–53].

2.3. Southern Blots and Riboprobes

BAC clones were digested with *SalI* and *NotI*, and fragments were separated by gel electrophoresis and transferred to a GeneScreen Plus hybridization membrane (Perkin-Elmer, Waltham, MA, USA) by capillary blotting [35,54]. The filter was evaluated with ³²P-riboprobes generated with RNA polymerases from linearized gene clones that served as templates to incorporate ³²P labeled ribonucleotides as described in [23,35,55]. The gene clones chosen for templates had an A6 element pattern (GenBank accession number EF607716.1), a B3 element pattern (EF607770.1), and a D1 element pattern (EF607784.1) [19]. After hybridization with the probes, the filter was exposed to X-ray film at −80 °C, which was processed with developer and fixer, scanned with epi-white light, and imaged with the Kodak Gel Logic 1500 Imaging System.

3. Results

3.1. Sea Urchin Genomic DNA Harboring the *SpTrf* Gene Family Is Unstable

Genomic instability is predicted for regions of DNA that contain many types of repeats including tightly linked genes with similar sequences [6,56], such as the *SpTrf* genes. An initial characterization of BAC DNA was carried out by *NotI* restriction digests, which released the insert from the pBACe3.6 vector to evaluate size. This approach identified 3% to 10% of the colonies from which BAC DNA was isolated after the 10-day growth period and had inserts that were smaller than expected when they included more than one *SpTrf* gene (Table 1). BAC-51-15 (see Table 1 for the BAC naming conventions) had a small insert compared to BAC-51-con (Figure 2A, blue vs. white arrows), and the corresponding gene amplicons indicated that the *SpTrf*-A2 gene was missing (Figure 2B, blue vs. white arrows). The other BACs that originated from BAC-51 all had full-length inserts and a full complement of genes (Figure 2A,B). BAC DNA isolated from *E. coli* colonies that contained

BAC-52 showed a variety of changes to the insert sizes (Figure 2C). BAC-52-19, BAC-52-2b, and BAC-52-4 all had small inserts compared to BAC-52-con (Figure 2C, colored arrows) and showed varying numbers of missing *SpTrf* genes (Figure 2D, colored arrows). BAC-52-19 did not support amplification of any *SpTrf* gene, indicating that all were missing (Figure 2D, red arrow). There was a single gene amplicon from BAC-52-2b, which indicated that *SpTrf-A2* was present, but the other genes were missing (Figure 2D, green arrow). BAC-52-4 (Figure 2C,D, yellow arrows), however, showed all gene amplicons, indicating that the full complement of *SpTrf* genes were present (Figure 2D, yellow arrow). Of additional note was BAC-52-4c, which had a full-length insert (Figure 2C, purple arrow), but none of the *SpTrf* genes were amplified (Figure 2D, purple arrow). Although the BACs with small inserts showed a variable loss of *SpTrf* genes, the majority of the deletions involved the *SpTrf* gene cluster. In contrast, BAC DNA isolated from multiple colonies that contained BAC-67 with only a single *SpTrf* gene showed no deletions (Table 1, Figure 2E), and the single *SpTrf* gene was maintained (Figure 2F). These results suggested that the repeats within the *SpTrf* cluster were associated with most of the DNA deletions, whereas a single gene separated from the rest of the cluster and with fewer repeats in the insert did not result in DNA deletions.

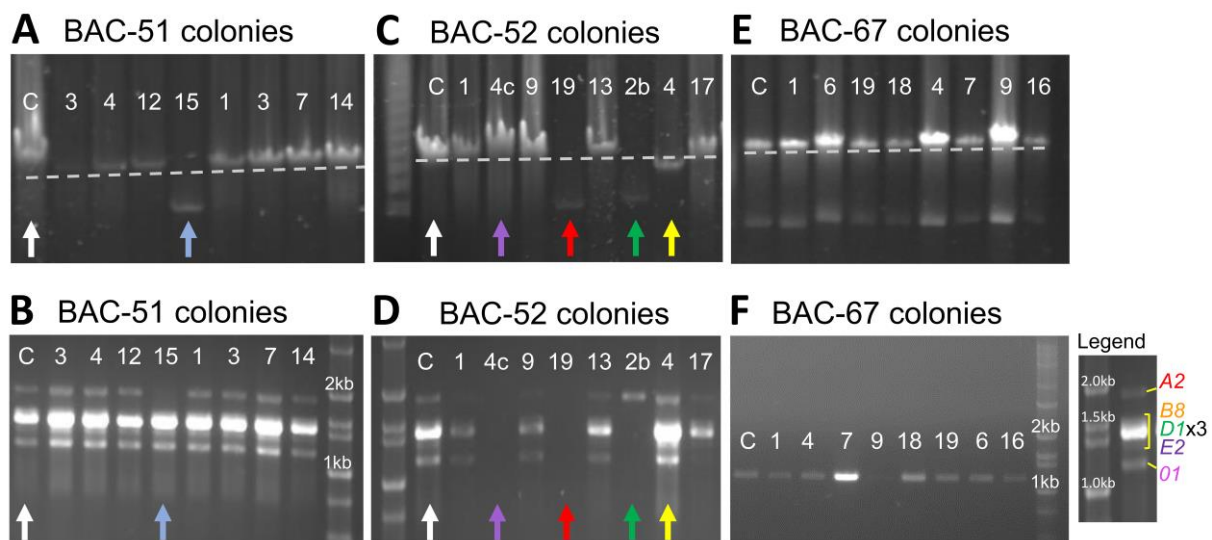


Figure 2. BACs isolated from some colonies after 10 days of growth have small inserts. (A,C,E) Representative *NotI* digests of four BACs containing *SpTrf* genes in locus 1, allele 1 identify BACs with decreased insert sizes. BAC-con clones (C lanes, white arrows) that were grown overnight once show full-length inserts. The C lanes are followed by eight representative samples of BAC DNA isolated from single colonies after 10 days of growth. The colored arrows indicate BACs with small inserts. The first lane in (C) shows a PFGE lambda ladder (Bio-Rad Laboratories, Hercules, CA, USA). (B,D,F) PCR amplicons of genes in BAC inserts illustrate the genes in the cluster and identify the genes that are missing after 10 days of growth. Genes were amplified using primers specific for *SpTrf* genes (see Materials and Methods). The colored arrows in (A,C) correspond to the arrows in (B,D), respectively. Colored arrows indicate the BACs in which the gene copy number is different from the BAC-con clones (white arrows). The first or last lanes of the gels show the DNA ladder (Hi Lo DNA Standard, Fisher Scientific, Hampton, NH, USA). The legend for *SpTrf* gene amplicons shows the bands that correlate with individual genes based on sizes. *SpTrf-A2* is the largest gene; the cluster of bands of intermediate size includes *SpTrf-B8*, *-D1*, and *-E2* (three *D1* genes result in the strongest bands in the center); and *SpTrf-O1* is the smallest. The marker is the Hi Lo DNA standard.

3.2. Deletions to the BAC Inserts Are Positioned in a Variety of Locations

Although the analysis of BAC insert size and gene copy number indicated a variety of DNA deletions, these results did not identify the locations of the deletions or whether there could be more than one deletion in an individual BAC insert. To address this question, the

BAC-con clones and three BAC clones with expected deletions were digested with either *XhoI*/*NotI* or *SacII*/*NotI*. To predict which fragments corresponded to regions of the BAC inserts to aid in the evaluation of the actual digests, virtual digests were used to evaluate full-length BAC insert sequences for BAC-51 and BAC-52 (Table 1). The virtual double digests generated a wide size distribution of bands and located the seven *SpTrf* genes in specific bands (Figure 3A). While most of the *SpTrf* genes were positioned on the same fragment because of their tight clustering (gene colors in Figure 3A,B are indicated in Figure 1), *SpTrf-A2* and *SpTrf-01* were more likely to be located on different fragments, which correlated with their distant locations at the edges of the cluster. These two genes have larger intergenic regions of 12 kb and 7.3 kb, respectively [35]. However, the *XhoI*/*NotI* digest for BAC-52 showed all genes on the same fragment except for *SpTrf-A2* (Figure 3A). The virtual digest results provided a framework for interpreting the fragments resulting from the actual digests.

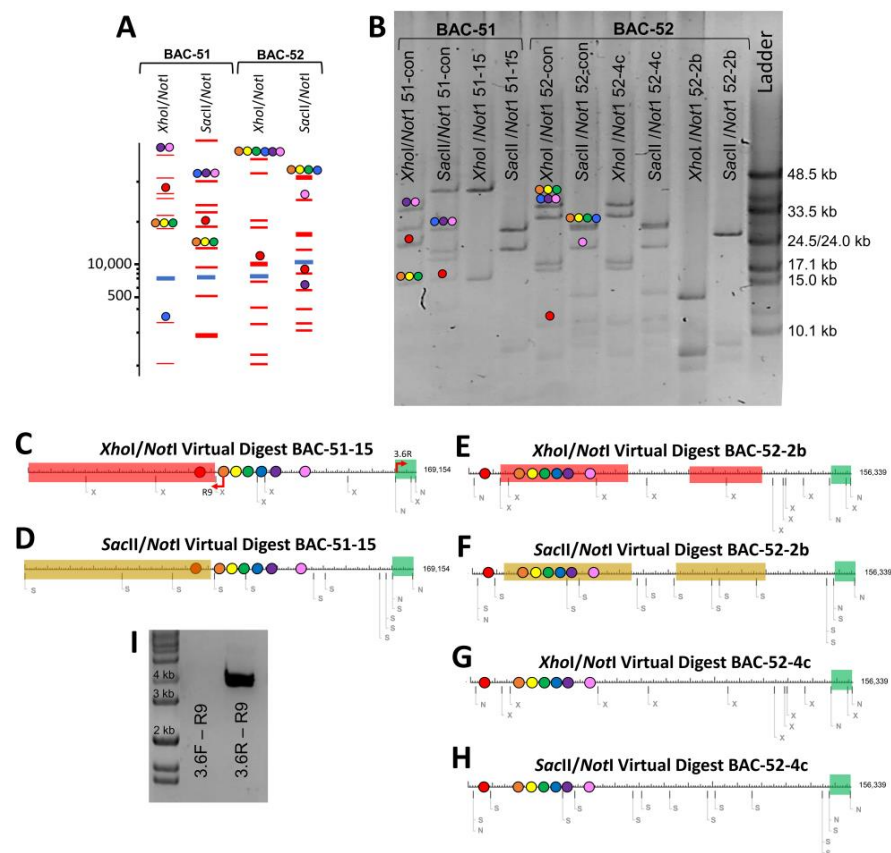


Figure 3. Locations of insert deletions are predicted by restriction digests. (A) *XhoI*/*NotI* and *SacII*/*NotI* virtual digests (<https://nc3.neb.com/NEBcutter/> accessed on 18 May 2018) of full-length BAC insert sequences show the fragment sizes and on which fragments the *SpTrf* genes are located. (B) *NotI* double digests with *XhoI* or *SacII* show altered band sizes from BAC clones with small inserts and multiple *SpTrf* genes. BAC-51-con and BAC-52-con with full-length inserts show bands of expected size based on the virtual digests. The ladder is lambda DNA (monocot λ mix; New England Bio-Labs). (C–H) Maps of virtual digests predict the locations of the deleted regions. The maps are shown in linear format, even though the BAC DNA is circular. The areas in red and yellow indicate the predicted deletions based on results in (B). The *SpTrf* genes are indicated by colored dots based on the gene colors in Figure 1. The green segment indicates the pBACe3.6 vector. Restriction endonuclease sites in the BAC DNA are indicated as N, *NotI*; X, *XhoI*; S, *SacII*. (I) PCR is used to orient and verify the size change in BAC-51-15 (see C). Primers (pBACe3.6F or pBACe3.6R) that surround the vector and the R9 primer that is located within each gene (see red arrows in (C)) confirm a ~90 kb deletion in BAC-51-15 that results in a 4 kb amplicon. The gel image is edited to delete irrelevant lanes and bring the DNA standard (Hi Lo; Fisher Scientific) next to the lanes of interest.

The actual digests with *NotI* and either *XhoI* or *SacII* of the DNA from BAC-51-con, BAC-52-con, and BACs with short inserts (Figure 3B) were compared to the virtual digests (Figure 3A). Differences were used to identify which bands were absent or had changed in size to deduce which regions of the BAC inserts had undergone deletions. Two of the three BACs showed evidence of deletions. The digests of BAC-51-15 indicated a large deletion in which most of the expected bands were missing (Figure 3B). The *XhoI/NotI* digest showed a loss of all but one band and a large change in size of another, which the *SacII/NotI* digest showed as a loss of all but three bands. When these deletions and size changes were mapped onto the virtual digest of BAC-51, results predicted a deletion of ~90 kb including the region expected to contain *SpTrf-A2* (Figure 3B–D). This verified the *SpTrf* gene amplicon results for BAC-51-15 in which the *SpTrf-A2* gene was missing (Figure 2D), thereby locating the position of the deleted region within the insert (Figure 3C,D). This deletion was also verified by PCR using the R9 primer (the annealing site is within the coding region of all *SpTrf* genes) and either the pBACe3.6F primer or the pBACe3.6R primer (annealing sites are at the ends of the vector; see red arrows in Figure 3C). An amplicon of 4 kb was generated from BAC-51-15 with pBACe3.6R, which was only feasible if a large deletion had brought the R9 annealing site in *SpTrf-B8* (orange dot) into close proximity to the vector (Figure 3C,I). Results for BAC-52-2b suggested two deletions, of which one removed all genes except for *SpTrf-A2* (Figure 3B,E,F). The second deletion in BAC-52-2b did not alter the *SpTrf* gene cluster; however, it indicated that multiple deletion events could occur in BAC inserts. BAC-52-4c was digested to determine whether any deletions were present given that it showed no change in insert size based on the *NotI* digest compared to the BAC-52-con (Figure 2C, purple arrow) and that it failed to amplify any of the *SpTrf* genes (Figure 2D, purple arrow). Results indicated that there were no differences in the digests for BAC-52-4c and the digests for the BAC-52-con (Figure 3G,H), suggesting that it had no deletions and that the absence of gene amplicons was likely a technical PCR failure (Figure 2D, purple arrow). These findings predicted that inserts that were smaller than expected for some BAC clones were the outcome of deletions. This also suggested that BAC inserts with multiple *SpTrf* genes that are associated with many types of repeats in the gene cluster [35,38] were the basis of the deletions. This was supported by results from BAC-67 with a single *SpTrf* gene and far fewer associated repeats, which did not undergo deletions (Figure 2E,F).

3.3. BAC Insert Sequencing and Assembly Verifies Gene Loss and Identifies the Edges of Deletions

To verify the results indicating BAC insert deletions based on changes in insert sizes and gene copy numbers, five BACs were sequenced (PacBio) and assembled into full-length insert sequences. BAC-51-15 and BAC-52-2b were selected based on predicted deletions, BAC-52-4c was selected because it maintained the full-length insert after the 10-day growth period but did not appear to have maintained the *SpTrf* genes (Figure 2C,D), and BAC-51-con and BAC-52-con were selected after growth for a single day. Notably, the preparation of large quantities of BAC DNA for sequencing required at least one and often more than one round of growth to acquire enough DNA of sequencing quality. The assembled insert sequences were aligned by hand to either BAC-51 or BAC-52 (Table 1) to verify the locations of the deletions (Supplementary File). Unexpectedly, all sequenced BACs had deletions, including BAC-51-con, BAC-52-con, and BAC-52-4c, which had been identified as full-length without deletions (Figure 2A,C; Table 2). Dot plots were used to illustrate the *SpTrf* gene cluster in BAC-51 and BAC-52 (Figure 4A,B) and to show the locations of deletions in the BACs that underwent the 10-day growth period (Figure 4C–G). The single deletion in BAC-51-15 was consistent with the size and location predicted by the virtual digests and included *SpTrf-A2* (Figure 4C). BAC-52-2b showed one large deletion that removed all of the *SpTrf* genes except for *SpTrf-A2* (Figure 4D) that was not consistent with the virtual digest predictions that suggested two smaller deletions (Figure 3E,F). It appeared that the two predicted deletions may have progressed to a single larger deletion that deleted the region between the two smaller deletions during the preparation for

sequencing. Surprisingly, during the preparation of DNA for BAC-51-con, BAC-52-con, and BAC-52-4c for sequencing, these BACs appeared to have also acquired deletions. The insert assembly and dot plot for BAC-51-con showed that, rather than a full-length insert with all *SpTrf* genes in the cluster, three deletions had occurred that truncated *SpTrf-A2* and *SpTrf-E2* and deleted *SpTrf-01* (Figure 4E). Similarly, the dot plot for BAC-52-con compared to BAC-52 also showed a deletion, which truncated *SpTrf-E2* and deleted five genes, although *SpTrf-01* remained (Figure 4F). Furthermore, dot-plot results for BAC-52-4c compared to BAC-52 showed that there were three deletions that removed about 100 kb, including *SpTrf-01*, and the other two deletions were located outside of the gene cluster (Figure 4G). This size change was not evident in the gel image of the *NotI* released insert, which appeared as the same size as BAC-52 (Figure 2C, purple vs. white arrows). The inconsistency within the results required further analysis to identify the source.

Table 2. The *SpTrf* gene complement is variable among BAC inserts.

BAC Clone Name ¹	Accession Number	Growth Period (Days)	BAC Insert Size; PFGE ² , Sequence	<i>SpTrf</i> Genes in Locus 1 Allele 1	Analyses ³ to Identify <i>SpTrf</i> Gene Complement
BAC-51	KU668451	na ⁴	Full-length 157,542 bp	All genes present	<i>NotI</i> digest Gene amplicons
BAC-51-con ⁵	na ⁶	1	Full-length 115,850 bp ⁷	All genes present	<i>NotI</i> digest Gene amplicons Virtual digests Insert sequence
BAC-51-15	PP082968	10	Short 67,180 bp ⁸	Deletion of A2 only	<i>NotI</i> digest Gene amplicons Virtual digests Insert sequence
BAC-52	KU668452	na ⁴	Full-length 144,728 bp	All genes present	Gene amplicons
BAC-52-con ⁵	na ⁶	1	Full-length 124,749 bp ⁷	All genes present	<i>NotI</i> digest Gene amplicons Virtual digests Insert sequence
BAC-52-4c	na ⁶	10	Full-length 76,739 bp ⁷	All genes deleted ⁹	<i>NotI</i> digest Gene amplicons Insert sequence
BAC-52-2b	PP082969	10	Short 36,374bp ⁸	All genes deleted except A2	<i>NotI</i> digest Gene amplicons Virtual digests Insert sequence
BAC-52-19	na	10	Short Not sequenced	All genes deleted	<i>NotI</i> digest Gene amplicons
BAC-52-4	na	10	Short Not sequenced	All genes present	<i>NotI</i> digest Gene amplicons Virtual digests

¹ Abbreviated BAC accession numbers are used as clone names. ² PFGE, pulsed field gel electrophoresis. ³ Virtual digests and gene amplicons are shown in Figures 2 and 3. ⁴ na, not applicable. The BAC insert was not sequenced for this study; it was acquired from GenBank. ⁵ con, control. BAC DNA isolated from a single colony grown for a single day served as the control. ⁶ The assembly artifacts precluded submission to GenBank. ⁷ The BAC insert length reported here includes deletion artifacts resulting from the assembly process. ⁸ Insert sequence and deletions are shown in the Supplementary File. ⁹ This is likely a technical PCR failure and a false negative (see Figure 2).

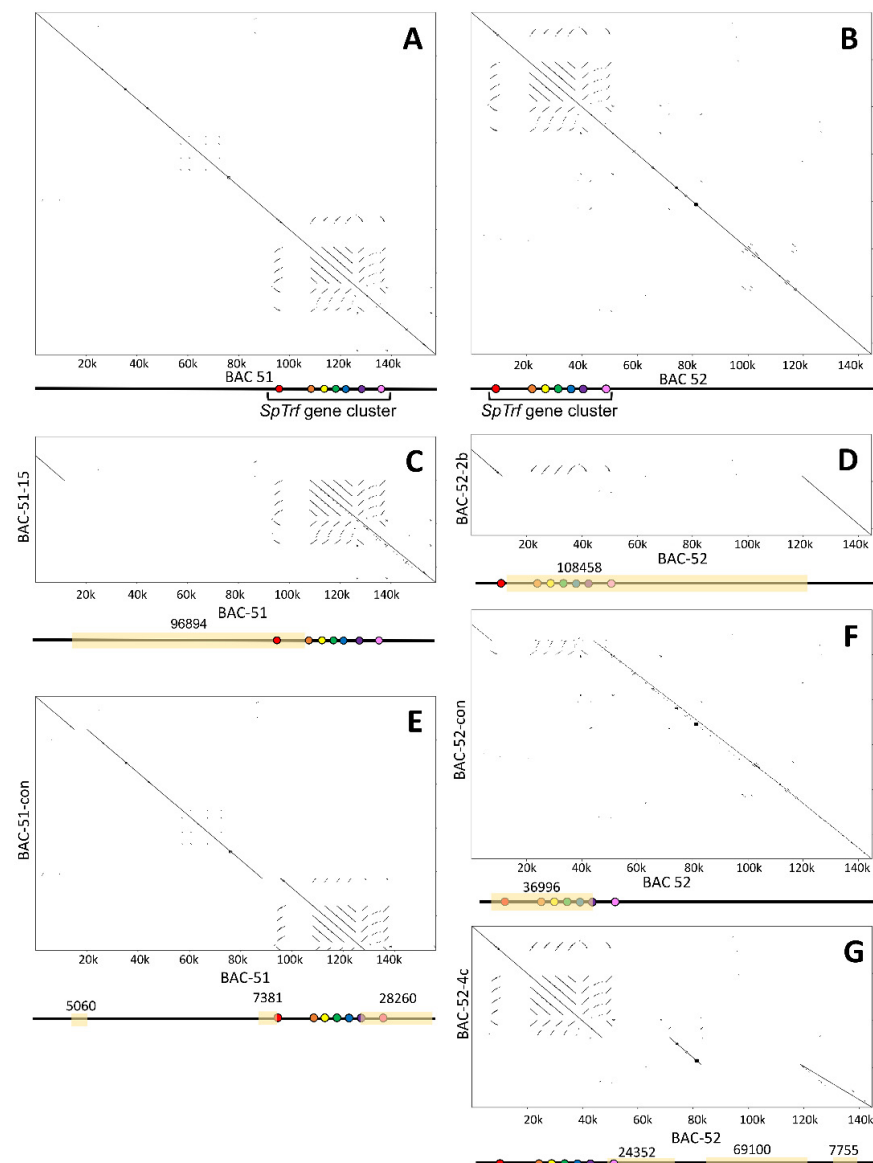


Figure 4. Dot plots of sequenced BAC inserts identify deletions by comparisons to the full-length BAC insert sequences. Dot plots compare each full-length BAC insert to itself and to other BAC inserts that show deletions. The gene cluster maps of BAC-51 or BAC-52 are shown at the bottom of each dot plot with the location of each gene indicated by colored dots (based on the gene colors in Figure 1). Deletions identified by the dot plots are indicated by yellow highlights in the cluster maps and the number of nucleotides in each deletion is indicated. (A) BAC-51 vs. self. (B) BAC-52 vs. self. (C) BAC-51 vs. BAC-51-15. (D) BAC-52 vs. BAC-52-2b. (E) BAC-51 vs. BAC-51-con. (F) BAC-52 vs. BAC-52-con. (G) BAC-52 vs. BAC-52-4c.

Because deletions in the full-length BAC inserts did not fit previous analyses of these inserts and because of reported difficulties in assembling sequence reads that include repeats [45,57], as we describe above, verification of the assemblies was required. Raw sequence reads from all five BACs were mapped against the reference sequences submitted to GenBank for BAC-51 and BAC-52 (see Table 1 for accession numbers). Results showed that BAC-51-15 and BAC-51-2b had deletions (Figure 5) in agreement with predictions from gene amplicons and both actual and virtual digests (Figures 2 and 3). BAC-51-con, BAC-52-con, and BAC-52-4c, which were predicted to be full-length with the full complement of genes (Figures 2 and 3), did not show deletions based on the mapping results (Figure 5), which was not in agreement with dot-plot comparisons of the assemblies (Figure 4). The

insert assemblies for BAC-51-con, BAC-52-con, and BAC-52-4c were likely the outcome of poor-quality sequence reads that the assembler program omitted when assembling the BAC insert sequences. Consequently, the deletions in these BACs were deemed to be assembly artifacts and illustrated the necessity to employ multiple means to verify sequence assemblies that include repeats. Indeed, the most common reason for low- or poor-quality sequence reads is the presence of highly repetitive stretches of sequences [57], which is the case for these BAC inserts that cover the *SpTrf* gene locus. Furthermore, poor-quality sequences in certain regions of the BAC sequences may be due to variability in deletion presence and location among the individual BAC clones isolated from individual bacterial cells that are present collectively in a single culture. The outcome for sequencing the BAC DNA with inserts that are unstable may be that some of the inserts are full-length and others have random deletions, leading to poor-quality sequence reads at specific locations. Overall, the BAC inserts with predicted and verified deletions showed that one or more *SpTrf* genes were removed.

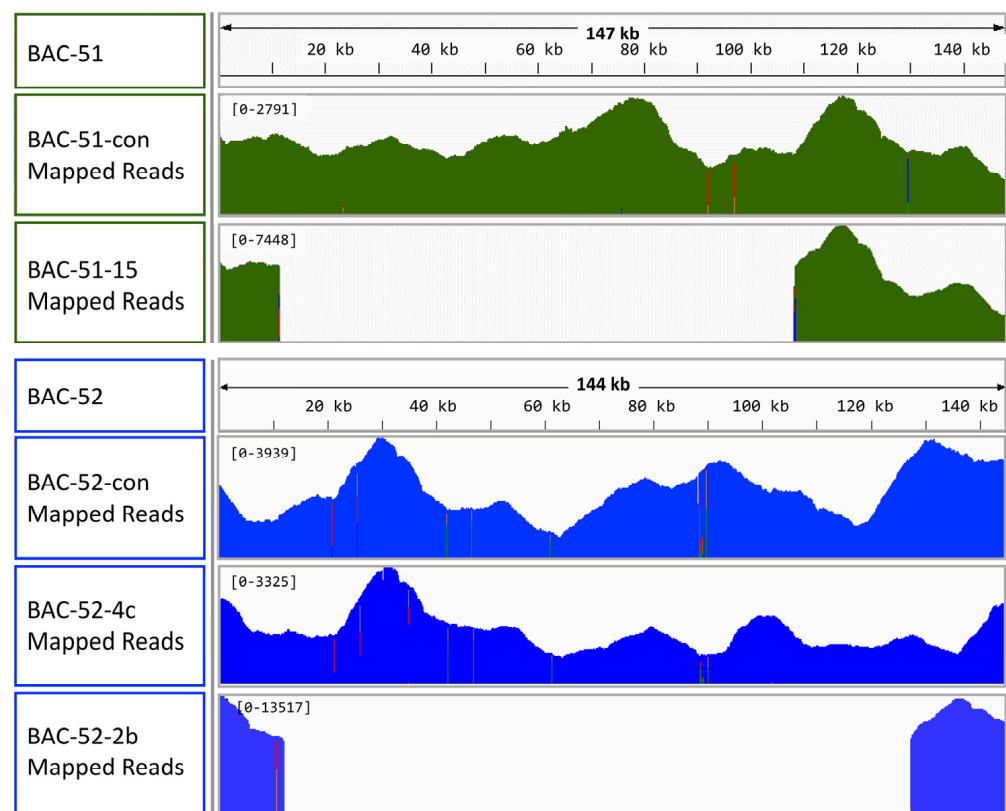


Figure 5. BAC insert assemblies generate false deletions from low-quality sequencing reads. Sequence reads for each BAC are mapped onto reference sequences for BAC-51 (green maps) or BAC-52 (blue maps) that have been reported previously [35]. The lengths of the reference sequences for BAC-51 and BAC-52 are indicated and a ruler is included for every 20 kb. Mapping histograms for each BAC compared to the reference sequence indicate the depth of sequencing coverage and reads per position from the raw PacBio sequencing data. Higher peaks indicate greater coverage, while the absence of peaks indicates no sequence coverage. The colored bars within the histograms indicate nucleotide positions that have increased nucleotide variability across sequence reads. The height of each colored bar indicates the number of sequences for a specific nucleotide. Bar colors indicate nucleotides: red (T), orange (G), light green (A), and dark blue (C).

3.4. BAC Deletions Are Flanked by STRs

Many reports of genomic instability and DNA deletions have focused on defects in DNA repair and replication or the locations of deletions and their associations with specific sequences, including STRs, other types of repeats, and poly G sequences that can form G-quadruplexes [58,59]. Consequently, the verified assemblies for BAC-51-15 and BAC-52-2b enabled an investigation of the sequences at the edges of the deletions. The two verified deletions in the BAC inserts identified from sequence alignments were located at or near regions containing STRs or polynucleotides. The first and larger deletion in BAC-51-15 was associated with CT STRs, which spanned 120 base pairs (bp) at the 5' end of the deletion and a region of CT and TA STRs of over 425 bp at the 3' end (Table 3; Supplementary File). The second and smaller deletion that was not verified by mapping results and was likely an assembly artifact based on poor-quality sequence reads (Figure 5) was not associated with repetitive sequences (Table 3; Supplementary File). BAC-52-2b had a single deletion, which was bracketed by poly G and poly C (poly G on the complementary strand; see Supplementary File) (Table 3). Poly G stretches presented the possibility of G-quadruplex formation that may impede DNA replication, leading to genomic instability [60]. In general, the locations of the verified deletions in the BAC inserts were associated with STRs and polynucleotides, in agreement with reports of DNA deletion associated with instability hotspots [6,58].

Table 3. Regions in BAC insert sequences that are the origins of deletions ¹.

BAC	Deletion	Local Sequence at Locations of Deletions ²
51 vs. 51-15	First, 5' end First, 3' end Second, 5' end Second, 3' end	CTCTCTCTCTCCCTTTCTCTCTCTCTCTCTCTCTCTCTCTCT//CCTCTACCTCTCTACTGTAT CTCTCACATTCTCCTTTTCTCTCTCTCTCTCTCTCTCT//ATCTAICTATACTCTACC CTCACAGTGTA AAAATGATTTCACAGTGTG//ACCAAAGTGTATAAAAGATT TTCACAGTGTG TTACAGTATACACAAGAAGTCATTCTCCCCGATTC//CATCATGCTTTGAACCTGTCACTCTCGTGA
52 vs. 52-2b	First, 5' end First, 3' end	TTTCTTTTTGCTCAACGGGGGGGGGG//TAGTGCATGCGCGGATCCAGGGAGGCCCGCCCCCCCCAAAA TACG CAGGATTTTTCAAGTGGGGGGGGGGGG//GGGTTTAAATTTTAAATCGGGCCGAAAATTTTCGCAT

¹ See the Supplementary File for the full-length sequence alignments. Bold font shows the locations of STRs or polynucleotide sequences. ² //, indicates the location of the deletion. Sequences to the 3' side of the deletion are present in the full-length BAC but are missing in the BACs that contain deletions.

3.5. Some BAC Inserts Are Deleted Prior to Analysis

The initial screen of the BAC library of sea urchin genomic DNA identified 75 BACs with *SpTrf* gene sequences; however, only 27 BAC clones supported PCR amplification of the *SpTrf* genes [35], of which we report results for BAC-51 and BAC-52 above. The clones that did not support amplification of the *SpTrf* genes were evaluated by Southern blots, and BAC-42 and BAC-44 were identified as containing *SpTrf* gene sequences (Figure 6). Based on these conflicting results, these BAC clones were submitted for long-read sequencing. The sequence reads for these BAC inserts could not be assembled into a single sequence and were instead assembled into eight contigs for BAC-44 and three contigs for BAC-42. No *SpTrf* gene sequences were identified within these assemblies, which contradicted the results in the Southern blot. The inconsistencies with BAC-42 and BAC-44 among the other BACs that failed to amplify *SpTrf* sequences suggested that the initial library screens had identified *SpTrf* gene sequences in 75 BAC inserts, but that during the growth and isolation of the BAC DNA for analysis by PFGE and PCR, the inserts underwent deletions. The inference was that these and the other 47 BAC clones were particularly unstable.

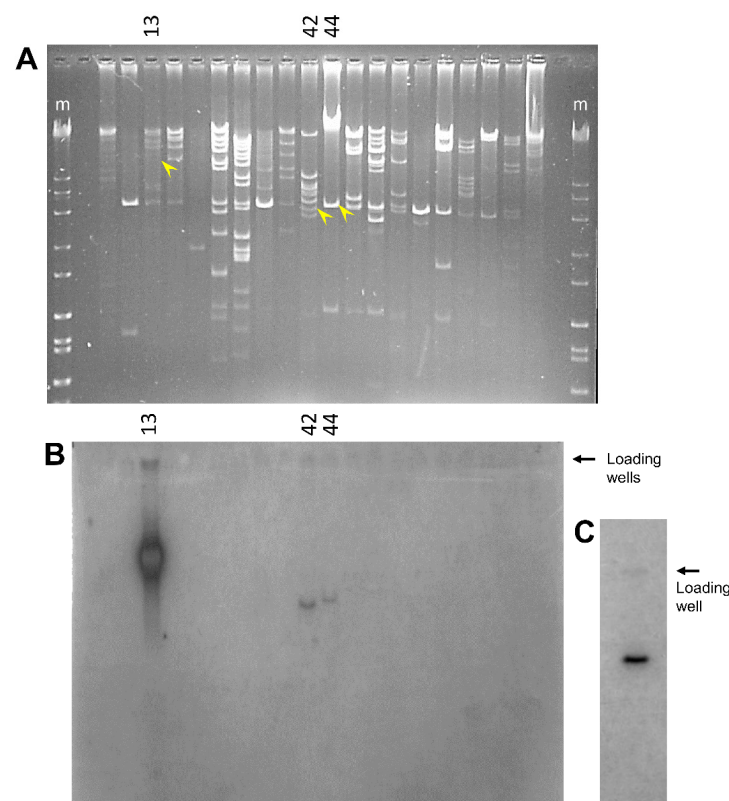


Figure 6. Many BAC inserts do not show *SpTrf* sequences by Southern blot. **(A)** BAC clones that did not amplify *SpTrf* sequences by PCR were digested with *SalI* and *NotI*, and the fragments were separated by gel electrophoresis, transferred to nylon filters, and evaluated with ^{32}P -riboprobes. A subset of those BAC clones are shown. The yellow arrowheads indicate bands with *SpTrf* sequences that correspond to the bands in **(B)**. The marker lanes (m) are Hi Lo DNA standards (Fisher Scientific). **(B)** The probes hybridize to three BAC inserts. The raw reads for BAC-42 and BAC-44 are available as BioSamples in GenBank, accession numbers SAMN39322606 and SAMN39322605, respectively. Preliminary sequence analysis of BAC-13 indicates that it includes a region of allele 2 for the *SpTrf* gene cluster in locus 1. Consequently, because this study focused on allele 1 of locus 1, long-read sequencing of the BAC-13 insert was not pursued. **(C)** BAC-7096 with six *SpTrf* genes (see Table 1, Figure 1B) [35,38] is the positive control for the Southern blot.

4. Discussion

4.1. Instability of BAC Inserts When Hosted by *E. coli*

Genomic instability can be lethal when it results in severe genomic fragmentation; however, genomic instability is also a source of potentially beneficial adaptations ([32,61]; reviewed in [62]). For the examples presented here, the BAC inserts are of no benefit to the bacteria; rather, it is the vector which contains the antibiotic resistance that is of benefit to the bacteria under the growth conditions in the presence of chloramphenicol. BAC inserts with repeats are unstable in prokaryotes ([63]; this study), and BACs with smaller inserts benefit *E. coli* because cells with smaller inserts can replicate the BAC DNA more quickly and with lower metabolic cost and therefore may proliferate faster than other cells with larger BAC inserts. The findings presented here suggest that BAC clones with multiple *SpTrf* genes are inherently unstable. Furthermore, deletions within the inserts may progress from multiple small deletions to larger deletions that incorporate the smaller deletions. This was likely the case for BAC-52-2b, with two small deletions identified in early analyses and a single deletion identified in the assembled insert sequence as one large deletion that incorporated the small deletions. Results suggest that the involvement of the STRs and polynucleotides, such as multiple Gs, may promote additional, larger deletions after the

initial smaller deletions are initiated. We propose that insert deletions in different BAC clones are initiated at different time points during the 10-day growth period. Perhaps early small deletions are more difficult to detect using our initial methods of restriction digests, whereas later, larger deletions in the same BAC are easier to detect. However, this pattern is not necessarily discernable from our dataset. We propose that the terminal condition of the BAC inserts is the deletion of all *SpTrf* genes, including the associated STRs and other types of repeats, as suggested from the insert sequences of BAC-42 and BAC-44 that do not include *SpTrf* sequences. Once the repeat sequences are deleted, subsequent deletions may cease to occur. Overall, the *SpTrf* gene cluster in BAC inserts appears to be very unstable and tends to undergo deletions. This instability has likely impacted the sequencing phase of the sea urchin genome assembly that employed the BAC library and was one aspect of the poor assembly of *SpTrf* gene loci of only 9 *SpTrf* genes, when 50 to 60 genes have been predicted in the family [33].

4.2. Genomic Instability of the *SpTrf* Gene Family May Underlie Variation in Gene Family Structure and Expression in Sea Urchin Cells

Although our findings suggest that the BAC inserts that include *SpTrf* gene clusters are unstable in *E. coli*, the key question is not about the growth benefits of smaller BACs for *E. coli* but whether local genomic instability applies to the *SpTrf* gene loci in the sea urchin genome. If the *SpTrf* gene family is unstable, and given the results presented here suggesting that the genes tend to be deleted from the BAC inserts, how might instability benefit the innate immune response of sea urchins? Genomic instability has been suggested as a source of beneficial genomic variation [62], and the STRs that surround genes and segmental duplications, the repeats within coding regions, and the sequence similarities among genes may all underlie gene duplications and/or deletions and changes in copy number plus subsequent sequence variations among genes [32,38]. STRs and other repeats have been the basis for proposed local genomic instability [4] and a theoretical evolutionary history of the *SpTrf* gene family [32]. Furthermore, there are three regions of GA STRs of several kilobases each that flank the genes in the *SpTrf* locus 2. These STR islands are located in positions that correspond to genes in locus 1, and this has led us to postulate that they may be the remnants of gene deletions that occurred in locus 2 [32,35].

Deletions and local genomic instability for the *SpTrf* gene clusters in the genome is in accord with previous reports on *SpTrf* gene expression and *SpTrf* gene copy numbers in single coelomocytes [29,42]. Immune challenge of sea urchins significantly increases *SpTrf* gene expression [23,24,64], *SpTrf* protein production [26], and the numbers of *SpTrf*⁺ coelomocytes in the CF and in other tissues [27,42,65]. Similar results for the *Trf* families have been reported for the sea urchins *Heliocidaris erythrogramma* [20] and *Paracentrotus lividus* [22]. It was assumed that individual phagocytes would express multiple *SpTrf* genes to drive swift responses to invading pathogens. However, when single phagocytes were evaluated for *SpTrf* gene expression, only identical *SpTrf* transcript sequences were identified for individual cells, implying expression of a single *SpTrf* gene per cell [29]. Although this result is consistent with gene regulation by promoters and enhancers, there may be an alternative explanation. In an approach to address the question of *SpTrf* gene regulation vs. gene deletion, single small phagocytes were sorted based on the *SpTrf* proteins on the surface, and red spherule cells that do not express *SpTrf* genes [29] but can be sorted based on the red color of echinochrome, which is characteristic of these cells [42]. Single sperm cells were employed as the control. Because the genes are small enough to be amplified by PCR, variations in amplicon sizes (see the legend in Figure 2) show that the arrays of genes are different among many of the coelomocytes, whereas they are all identical for each sperm for a given animal [42]. Furthermore, the *SpTrf* gene copy number is reduced in most coelomocytes compared to sperm. When the results for *SpTrf* gene expression and gene copy number are integrated, the interpretation suggests that the coelomocytes alter the *SpTrf* gene family, which may restrict and enhance the expression of a single gene per cell. These results are consistent with the notion of local

genomic instability of the *SpTrf* gene family and the hypothesis that putative DNA repair mechanisms are employed by the coelomocytes to balance control of locus instability with enhancing *SpTrf* gene sequence diversification. The outcome would be to the advantage of sea urchins in the arms race between their innate immune system and potential pathogens. Parallel results of local genomic instability have been reported for the *agglutinin-like sequence* multi-gene family in *Candida albicans* that contain tandem repeats and encode variations in adhesion proteins for binding to and colonizing host endothelial and epithelial cells, which is an example of a pathogen–host arms race ([66]; reviewed in [62]). Similarly, genes in the fungal *hmv* family, which functions in allorecognition specificity, encode a series of WD40 repeats that form β propeller structures and show local instability, driving variations in the numbers and sequences of repeats and altering interactions with conspecifics [67]. The killer immunoglobulin-like receptor (*KIR*) locus functions in natural killer cells (reviewed by [68]) and displays gene sequence diversity, tight gene clustering of less than 3 kb of intergenic space, and extraordinarily fast evolutionary change to allelic sequences and the locus structure, as suggested by extensive crossing over [69,70]. The range of repeats in the *KIR* locus is consistent with local genomic instability that drives the variability [71]. The first intron of the *KIR* genes is a minirepeat composed entirely of 30–60 repeats of 19–20 bp [71], and the locus has a number of transposable elements [13], which are repeats that may also result in *KIR* locus instability. Hence, the vertebrate *KIR* and the sea urchin *SpTrf* families both have extensive repeats that may be involved in their respective sequence diversification that may be beneficial in the host–pathogen arms race [72].

Hotspots of genomic instability that are often associated with STRs result in slow or poor DNA replication due to replication fork stalling or reversal, leading to double-strand DNA breaks [73,74]. Non-B DNA structures, such as Z-DNA, hairpins from inverted repeats, and poly G stretches that may form G-quadruplexes can also be the basis for genomic instability at hotspots (reviewed in [75]). G-quadruplexes can block DNA replication fork progression, leading to local genomic instability when helicases fail to unwind G quadruplexes in eukaryotes ([76,77]; reviewed in [78]) and in *E. coli* [79,80]. We report both possibilities for verified DNA deletions in the BAC inserts; CT STRs are associated with the deletion site in BAC-51-15, and poly G regions are associated with the deletion site in BAC-51-2b. In general, the wide variety of DNA repeats that are associated with the *SpTrf* gene family has suggested local genomic instability [32,35,38]. The structural appearance and organization of the family is consistent with instability of the genes located in segmental duplications, STRs that flank both genes and segmental duplications, tandem and interspersed repeats in the coding sequences, sequence variations among the genes, and proposed gene deletions in locus 2.

DNA damage resulting in local genomic instability includes (i) DNA replication stress at fragile sites composed of STRs and other types of repeats [6], (ii) R loops of DNA–RNA hybrids that can lead to double-strand breaks [81,82] and double-strand breaks that occur during general DNA replication (reviewed in [83,84], (iii) highly expressed genomic regions resulting in DNA damage from transcription–replication conflicts [85,86], and (vi) DNA tangles resolved by cleavage followed by incorrect re-ligation [87]. DNA damage and genomic instability are counteracted by DNA repair mechanisms and expression in response to DNA damage in eukaryotes [88]. Genomic instability of the *SpTrf* gene clusters that could result in the deletion of entire loci must be regulated in some way, given that the gene family is maintained in genomes of euechinoid species. Molecular control by coelomocytes to promote, block, or repair chromosomal changes in genomic DNA regions of the *SpTrf* gene clusters may correlate with variations in the level of expression for the DNA repair mechanisms. Hence, there may be a correlation between the expression level of the *SpTrf* genes, which are upregulated significantly in response to immune challenge (reviewed in [89]), and the sea urchin DNA repair mechanisms that may control or regulate local instability of the *SpTrf* gene clusters. Because the *SpTrf* gene family shows an identical composition among single sperm cells from individual sea urchins [42] and given that DNA damage is associated with DNA replication, DNA repair genes may be expected to show

elevated expression during mitosis and meiosis in gonads to maintain the structure and membership of the *SpTrf* gene family. Conversely, because individual coelomocytes do not appear to maintain the *SpTrf* gene family equally among cells, DNA repair gene expression may be reduced and/or variable among cells in the axial organ and pharynx where the coelomocytes proliferate [27]. DNA repair genes were identified in initial annotations of the *S. purpuratus* genome sequence (version 2.1) [90], although their expression has not been investigated for the tissues and organs of adult *S. purpuratus*. However, gene expression related to DNA repair has been documented for sea urchin larvae and coelomocytes responding to exposure to genotoxic chemicals [91]. Searches of the *S. purpuratus* genome sequence (version 5.0; www.Echinobase.org; [36], as of 15 December, 2023) result in a number of genes that encode proteins with putative DNA repair functions, including general DNA repair, DNA repair and recombination, excision repair, mismatch repair, double-strand break repair, and DNA cross-link repair, in addition to exonuclease and helicase functions (see also Table S2 in [90]).

Local genomic instability in the *S. purpuratus* genome may not be random, based on the findings from BAC clone insert instability in *E. coli*, yet sea urchin cells must have a means to control or regulate instability to take advantage of the characteristics of the *SpTrf* gene clusters, which are riddled with a wide range of repeats with predicted locations of duplications, deletions, and insertions [32,38,42]. In the case of the *SpTrf* genes in euechinoids, we propose that local genomic instability is an initial and required parameter to drive gene sequence diversification in the population that results in an immune response gene family that keeps pace in the arms race with the marine microbes with which sea urchins share their habitat.

5. Conclusions

Genomic instability is driven by the presence of repeats in local regions of the genome. The *SpTrf* gene family, which functions in immune response in the purple sea urchin, is surrounded by multiple types, sizes, and numbers of repetitive sequences, including flanking STRs. We show that the inserts of BAC clones with multiple *SpTrf* genes are unstable in *E. coli* and that with continued growth over time, one or more of the *SpTrf* genes are deleted. The deleted regions are commonly bracketed by STRs or polyG stretches. These results suggest that the repeat-riddled region in the sea urchin genome that includes the *SpTrf* gene family is locally unstable. This may result in expanding, contracting, or maintaining members of the *SpTrf* gene family, which is likely to be beneficial in the arms race against pathogens.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/genes15020222/s1>. Supplementary File: BAC insert sequence alignments.

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Data Availability Statement: The sequence data generated in this research are available from GenBank. See Tables 1 and 2 for accession numbers. The raw sequence reads for BAC-42 and BAC-44 are available from GenBank (BioSample accession numbers SAMN39322606 and SAMN39322605, respectively). The GenBank accession numbers for BAC-51-15 is PP082968, and BAC-52-2b is PP082969. The BAC clones used in this analysis and their genomic DNA library locations are as follows: BAC-51 location is 10B1; BAC-52 location is 4074J14; BAC-67 location is 14K16; BAC-42 location is 4069G2; BAC-44 location is 4069C2, and BAC-13 location is 3020I13. See also: <https://www.echinobase.org/echinobase/>.

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Local genomic instability of the SpTransformer gene family in the purple sea urchin inferred from BAC insert deletions

Megan A. Barela Hudgell, Farhana Momtaz, Abiha Jafri, Max A. Alekseyev, L. Courtney Smith

Methods

The sequence reads for each BAC insert were assembled into contigs as described in the main paper. The consensus sequences were aligned by hand in Molecular Evolutionary Genetics Analysis X (MEGAX) against the original BAC insert sequences (GenBank Accession numbers KU668451 [BAC-51], KU668452 [BAC-52]) (1). The alignments shown below were generated in BioEdit (version 5.0.9), formatted and exported as rtf files, and imported into this document.

BAC insert sequence alignments

Alignment 1. BAC-51 and BAC-51-15 identify the locations of three deletions in BAC-51-15

Page

1

Deletion 1; 96,854 bp

Deletion 2; 2,270 bp*

* likely represents an assembly artifact

Alignment 2. BAC-52 and BAC-52-2b identify the locations of one deletion in BAC-52-2b

9

Deletion 1; 108,397 bp

Alignment 1. BAC-51 and BAC-51-15

	18910	18920	18930	18940	18950	18960	18970	18980	18990	19000	
BAC-51										
BAC-51-15	TCATTCAACATACCGTAAATAGCTTCATCATGTTCTTTT	CAGAAGATTTTAATTTAAAATATTTT	TATTGTGGTTGGGGTATGTTTTCTCCA	ACTGCAAATG							
	19010	19020	19030	19040	19050	19060	19070	19080	19090	19100	
BAC-51										
BAC-51-15	ATGTGCATTATATTTTATAAGGTTTCATTTTAAATAAGAACTGTAATAACCTAGTATTTTTGATCAAATATGTTGCTCAGAAGTCTTGAACAGCATTAA										

[illegible]

Deletion 1, 5' end

2010 2020 2030 2040 2050 2060 2070 2080 2090 20300
|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|
 BAC-51 CTGTATGTGTCTGTTTATCTCTCTACAGATGTGGTAGAGTTTAGGGGTCAGTTTACACTGAAGAGCATTGAGAATCCTGGAACCGTTCTGTCTGAATACC
 BAC-51-15 -----

20310 20320 20330 20340 20350 20360 20370 20380 20390 20400
 BAC-51 AATCATGCTGCTCAGATCCATCCAGCAGTACCTACCAAGCACTTCAGAACAGAATCTATGATCATGTAAGTATACCAAGATAGTTATATATAGTCAAATT
 BAC-51-15 -----

Sequences (96,854 bp) in BAC-51 that are deleted in BAC-51-15 are omitted

116610 116620 116630 116640 116650 116660 116670 116680 116690 116700

BAC-51 GAGAGAGAGAGAGAGATCCTGCTACTACTGTCGCTAGATTACTGCTATAAAAACTACTATTTCTTAAAGGCAAGTACACTCCAAAAATACCTTACT

BAC-51-15 -----

	116710	116720	116730	116740	116750	116760	116770	116780	116790	116800
									
BAC-51	TTCAATAGAAATCAGACAATATGTAATCAGACAATTATTTCCCTCACTTTCATTATATATAAAATTTCTTTGTCTATCTTCTTCTCTCTCTCCCC									
BAC-51-15	-----									

[illegible]

116910 116920 116930 116940 116950 116960 116970 116980 116990 117000
|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|....|
 BAC-51 TGTCTTTGTCTCTTTGTCTCTCCCTCCCCCCCCCTCTCTCTCTCTCTATATATAATCAGACAATATATAATACCTATATCCCTCACTCTCACTTTTCTC
 BAC-51-15 -----

[illegible]

Deletion 1, 3' end

[illegible]

	118210	118220	118230	118240	118250	118260	118270	118280	118290	118300
BAC-51									
BAC-51-15	TCCTTGCCCGAAGAAAGGTCTTCCTCCAGCACCATCTGGTCTGGAGCCACCAAATCCAGGTCCGTGCAATCTCCTACCACCCATCGGACCGCCATTTTGT									
	TCCTTGCCCGAAGAAAGGTCTTCCTCCAGCACCATCTGGTCTGGAGCCACCAAATCCAGGTCCGTGCAATCTCCTACCACCCATCGGACCGCCATTTTGT									

	118310	118320	118330	118340	118350	118360	118370	118380	118390	118400
BAC-51									
BAC-51-15	CTGCGTCCATCCATATGTGGGGCACCAAATCCAGGTCCATCGAACCTCCTTCCACCCATTGGTCCACCATCTTGCCTAGATCCACCCATCTGCATTCCAC									
	CTGCGTCCATCCATATGTGGGGCACCAAATCCAGGTCCATCGAACCTCCTTCCACCCATTGGTCCACCATCTTGCCTAGATCCACCCATCTGCATTCCAC									

	118410	118420	118430	118440	118450	118460	118470	118480	118490	118500
BAC-51									
BAC-51-15	CAGGCCTTCCTCCAAAGCGACCTTGTCTCTCTCTGCCATTCTCATTTCTCGCCGTTTCATTGAAATCTCTTCGTGCGTGAGCTGTAAGGGTTAATGA									
	CAGGCCTTCCTCCAAAGCGACCTTGTCTCTCTCTGCCATTCTCATTTCTCGCCGTTTCATTGAAATCTCTTCGTGCGTGAGCTGTAAGGGTTAATGA									

	118510	118520	118530	118540	118550	118560	118570	118580	118590	118600
BAC-51									
BAC-51-15	AATAATCGAATTAATTAAGTAATCACATTCTGCATTATTATATTTGGCTCTACATTGGTTTATAGCAGGTGACAAATAAGAATGGTAACTCGCCCTTCTTA									
	AATAATCGAATTAATTAAGTAATCACATTCTGCATTATTATATTTGGCTCTACATTGGTTTATAGCAGGTGACAAATAAGAATGGTAACTCGCCCTTCTTA									

	118610	118620	118630	118640	118650	118660	118670	118680	118690	118700
BAC-51									
BAC-51-15	TATTTTTT-TCCCTTTATGTTTAATCCTAATCCGTAAGTGCGGTTTCGATATTCAAACGCGCTAATTCTGACTGAATTAAATTTGATTACCAGATTGAACGT									
	TATTTTTTTCCCTTTATGTTTAATCCTAATCCGTAAGTGCGGTTTCGATATTCAAACGCGCTAATTCTGACTGAATTAAATTTGATTACCAGATTGAACGT									

	118710	118720	118730	118740	118750	118760	118770	118780	118790	118800
BAC-51									
BAC-51-15	AAACATAATACATCCTTTGTCATTTCAGAAAACGGATTGGTGTAGGCTTAATAAATACAATTAATTATATTGTGTAACGAACAATTTAAAAATGCATAT									
	AAACATAATACATCCTTTGTCATTTCAGAAAACGGATTGGTGTAGGCTTAATAAATACAATTAATTATATTGTGTAACGAACAATTTAAAAATGCATAT									

	118810	118820	118830	118840	118850	118860	118870	118880	118890	118900
BAC-51									
BAC-51-15	TACAAACTATGAAATAATCATATTTACTTCTGTGAGCCCTTCGTTGGCTTTATATTTAGCACTTATCAAGTAATACCGAGTAATAATTTGATTTCTTACC									
	TACAAACTATGAAATAATCATATTTACTTCTGTGAGCCCTTCGTTGGCTTTATATTTAGCACTTATCAAGTAATACCGAGTAATAATTTGATTTCTTACC									

	118910	118920	118930	118940	118950	118960	118970	118980	118990	119000
BAC-51									
BAC-51-15	CGAGATAGCAAGAGCAGCCACAATGGCAACGATCAATGTTGCTTTACCTCCATGTTTGTAAGGTCTCTCCGATGCTACAAGCTTTCTCTAGATTTCGTTG									
	CGAGATAGCAAGAGCAGCCACAATGGCAACGATCAATGTTGCTTTACCTCCATGTTTGTAAGGTCTCTCCGATGCTACAAGCTTTCTCTAGATTTCGTTG									

	119010	119020	119030	119040	119050	119060	119070	119080	119090	119100
BAC-51									
BAC-51-15	CCTTCCAAGAGAGAAGTACCTCCAACTTAACAACCTACCTGCTGGGCCACTGAATTTATAGGTTTTCTACCTAGATTGATATCTCACACTAGTACCAGA									
	CCTTCCAAGAGAGAAGTACCTCCAACTTAACAACCTACCTGCTGGGCCACTGAATTTATAGGTTTTCTACCTAGATTGATATCTCACACTAGTACCAGA									

Matched sequences (37,500 bp) between the BACs are omitted

	156610	156620	156630	156640	156650	156660	156670	156680	156690	156700
BAC-51									
BAC-51-15	GGGCACCTTATCAGAGAAGAAAAGGAGTAGTTATAAAAAGGAG--AAGGGGCACTCTGTTTTAATGAAAAGCGCATTTATTAGAAAAGGTAAAGGGGCACT									
	GGGCACCTTATCAGAGAAGAAAAGGAGTAGTTATAAAAAG-AGAAGAAGGGGCACTCTGTTTTAATGAAAAGCGCATTTATTAGAAAAGGTAAAGGGGCACT									

	156710	156720	156730	156740	156750	156760	156770	156780	156790	156800
BAC-51									
BAC-51-15	TGTCAAAGTAAACGAGCATCTCTCATATTTGAATAGGGGCAATTATCGGACGTGAAAGAAGCACTAGTCTTATCTGAAATGAAAAGGAGCAACTTCTTT									
	156810	156820	156830	156840	156850	156860	156870	156880	156890	156900
BAC-51									
BAC-51-15	TTACCTGAAAGGGGTCATTACCAGAGCTGAAAAGAGGCATTTATGAAAAGGAGAAGGGGATCTTTGTTTTAATGGGG-AACTAACCATCAGGAAAAGGG									
	156910	156920	156930	156940	156950	156960	156970	156980	156990	157000
BAC-51									
BAC-51-15	GCACCTCTCAAATGTTAATAAGGCAATTATCAGAGGTGAAGGGGGCACTTATTACAAGTGAAGGGCGCACGTATCGTACCTGAAAAGAGGCACCTTACTA									
	157010	157020	157030	157040	157050	157060	157070	157080	157090	157100
BAC-51									
BAC-51-15	GACATTAAAAGAGGCACACAGCATGAGTAGTAAACACAGCCGTGGAAAAATGTAAGCAACACAAGTTCGTATATAGTCGTACGATGAGCGATTGAAACAA									
	157110	157120	157130	157140	157150	157160	157170	157180	157190	157200
BAC-51									
BAC-51-15	CGAAGTCTGACGGATCTTTGGTCTGAAGAGCACACCTGTCTCAGCACTGGCGTACTGGTAGGGGGCTAGGGG-AGGGGGGGGGG-CGCCATCCCCCAAATAAA									
	157210	157220	157230	157240	157250	157260	157270	157280	157290	157300
BAC-51									
BAC-51-15	ATTGAACCATGACATATTTTCAATTCATATACAAGTGATGTATAAATGTTTCGTGATATCATATGATTTTCAACAAAATGCGTTTTTCTCCATTTCTGTT									
	157310	157320	157330	157340	157350	157360	157370	157380	157390	157400
BAC-51									
BAC-51-15	GCAAACCTGACACATTCCCGAAGCCAGGAAGAGCATATGTTTTTTTTTATAATCACTAAGGGGCTTATAGTGGCCACTAACAGTAACACTTGTATAAAAAA									
	157410	157420	157430	157440	157450	157460	157470	157480	157490	157500
BAC-51									
BAC-51-15	ACACACTCATTACAGTGTGAACACACAGTTCACACTGTGAACGGAGCCTTTTCACAGTGTGGACACTGTCACAATGTGGTAAA--CCACACAGTATTCA									
	157510	157520	157530	157540	157550	157560	157570	157580	157590	157600
BAC-51									
BAC-51-15	CATTGTAAAATTCGATTTTCACAGTGTGAATGTACCTGCTCACAGTGTGAAAATGATTTTCACAGTGTGACCAAACCTGTGATAAATAGATTTTCACAGTGTG									

Deletion 2, 5' end, this is likely an assembly artifact

	157610	157620	157630	157640	157650	157660	157670	157680	157690	157700
BAC-51									
BAC-51-15	AATACCTTACGACATACATTTACGGTGTGAACAGTAAATTTCTCAGTGTGGCCACTCTGTGAAAAATACTTCACATTGTGAAAAATAGAAATTCACAGT									

	157710	157720	157730	157740	157750	157760	157770	157780	157790	157800
BAC-51									
BAC-51-15	ATGGCCGCTGGAATACTAGTGTTTTGGATCGAGGCCAATCAAGTTTCCGTGATAACAGACGAATATGTACAACTACATGTTGACGGATGAGAACATAATT									

	157810	157820	157830	157840	157850	157860	157870	157880	157890	157900
BAC-51									
BAC-51-15	GGAATGGCGGACATGGCCTGGTGAAGTTTCCCAACGGCTAGGATAGCTATCAGGCGAGGGTGTTGCCGGGCTGCTGCCGGGCTGTCATTGAATGGTGAGT									

Sequence (2,270 bp) in BAC-51 that deleted from BAC-51-15 is omitted, this is likely an assembly artifact

	159510	159520	159530	159540	159550	159560	159570	159580	159590	159600
BAC-51									
BAC-51-15	TCCTGGAGTATGCGTGCGTGACAGTGCGGGATCCTCACACCCAGGACAACATACACAGGTGGAAATGGTACAGCTCCGCAACGCCAGTTTATCACTG									

	159610	159620	159630	159640	159650	159660	159670	159680	159690	159700
BAC-51									
BAC-51-15	GAGATCACCATCATAACCAGCAGCATCGCACCAATGCTGCAACATCTCAAATGGCCATCTCTCCGAGAACGCAGGGCACGGTTTAAGATGGTGATAGTGTA									

	159710	159720	159730	159740	159750	159760	159770	159780	159790	159800
BAC-51									
BAC-51-15	CGGAACTGTCAACCTGCACTCAGAGTCTTCTCAAAATGTACCTTCTATCGGTGTTAACATCTACCATGGTCATACCCAGTAATTCCAAATACAATTTACA									

	159810	159820	159830	159840	159850	159860	159870	159880	159890	159900
BAC-51									
BAC-51-15	ATGACTCTCGTATACCAGAAGTCATTCTTCCCGATTCCATCATGCTTTGGAACTTGCTACCTGCTGGATTGTCAACTGTACTTTTCGCTAAAGCTTTTA									
	-----CATCATGCTTTGGAACTTGCTACCTGCTGGATTGTCAACTGTACTTTTCGCTAAAGCTTTTA									

Deletion 2, 3' end, this is likely an assembly artifact

	159910	159920	159930	159940	159950	159960	159970	159980	159990	160000
BAC-51									
BAC-51-15	AGCAAGAGGTACAAAATTTCCATCTCCGTTAGAAGGGGAAGAAGGATTTTAACTGCTCATAGAATATTATAGAAAGTTTAAATTCGCACTGTATATTC									
	AGCAAGAGGTACAAAATTTCCATCTCCGTTAGAAGGGGAAGAAGGATTTTAACTGCTCATAGAATATTATAGAAAGTTTAAATTCGCACTGTATATTC									

	160010	160020	160030	160040	160050	160060	160070	160080	160090	160100
BAC-51									
BAC-51-15	GCGACACCAGACCTGACGTACGTGATACACCATACGGTGGACTGGAGTGATATCTCAAGATCTGGGGGATGATAATTCCGGGCTTTAACAAATTTGTGTGAT									
	GCGACACCAGACCTGACGTACGTGATACACCATACGGTGGACTGGAGTGATATCTCAAGATCTGGGGGATGATAATTCCGGGCTTTAACAAATTTGTGTGAT									

	160110	160120	160130	160140	160150	160160	160170	160180	160190	160200
BAC-51									
BAC-51-15	AGTGGAGGTATTATTTTTGACTGCACAGTCGGAAGTTCCATCTAGTTATAATATCCTAACATCAGTATATTATCGCTAGAACAAAAAATCATAAGTTACT									
	AGTGGAGGTATTATTTTTGACTGCACAGTCGGAAGTTCCATCTAGTTATAATATCCTAACATCAGTATATTATCGCTAGAACAAAAAATCATAAGTTACT									

	160210	160220	160230	160240	160250	160260	160270	160280	160290	160300
BAC-51									
BAC-51-15	TATGTAGTTTAGGAAGTACATATATTTTTGTATGCCATTCTGCTTTTTTCTGTGCCACCGTATATAGAGTGTATGGTTATTTATTTGTAAAAAGATGAGC									
	160310	160320	160330	160340	160350	160360	160370	160380	160390	160400
BAC-51									
BAC-51-15	AAAATCTGTTTTTTTCTCATCTTCAGGTATTTATTAGTATTAGTTGTAGAAATTTATGTAGCCATAACAGTAAAACGTGTCTGCCTAAAATAGATTGT									
	160410	160420	160430	160440	160450	160460	160470	160480	160490	160500
BAC-51									
BAC-51-15	TTTCAACTTTTTTTTCAAGATGTATTGACAGAGATATTTTGAACCTTTTTATCATTTTAATAGGTTTATGAAGAACCTGTATGTGCCAAAGATTTTCTGAT									
	160510	160520	160530	160540	160550	160560	160570	160580	160590	160600
BAC-51									
BAC-51-15	ACATTGCAATTGCCAAAGTTACATGTCATGTGTGGATATTTCTATACTTGGAAATGTGACTAGCCATACTAATGTATCTTCATAAATTTAATATACCTAC									
	160610	160620	160630	160640	160650	160660	160670	160680	160690	160700
BAC-51									
BAC-51-15	AGAAATGTTTAAAGAGATATTGCCAGAGATGCTCTAAACCTATTTT-ATTACTGACGTAACATGCTAATGTGCTTTTATTATGTTTTACGAGTGTGAGTAC									
	160710	160720	160730	160740	160750	160760	160770	160780	160790	160800
BAC-51									
BAC-51-15	GAGTTCAGGGAAATATTGATGATATGTGCAATTTTTGCCGTCACCTGTAATTTATTCCTCGATGTTATACAGTGGAAACCTCTATAACTCTATTTCACAAGT									
	160810	160820	160830	160840	160850	160860	160870	160880	160890	160900
BAC-51									
BAC-51-15	CCACGTTTTTCTGTAATTCATTGTTTTAAACCCTTGTACAATAAGATATCTGATATAAAGAGGTCAATTTAGTGGACACCAATGTGCTCATTATAATGAG									
	160910	160920	160930	160940	160950	160960	160970	160980	160990	161000
BAC-51									
BAC-51-15	GTTTCACTGTAGATAGCAAGTTCTGTTTTTCAAGATATCTCTACTATCTAATTTAAACTGTTTGGATTCAATTTGTCTTTGTTTTCACTCTGGTTCACATC									
	161010	161020	161030	161040	161050	161060	161070	161080	161090	161100
BAC-51									
BAC-51-15	ATGTTGGCATTACAGAGTAAAATTTGTTTATCTGTAAGTATAGTATTACTGCAATCTTTGTATACCTAAGAGGTATATGTTTCATAAGTCCTTTTAGCGA									
	161110	161120	161130	161140	161150	161160	161170	161180	161190	161200
BAC-51									
BAC-51-15	AAAGTACATCAATATAAGCTTTTGAATACATTTTAAGTTACCTCATAAAGGCTATAACATGTATATGTTTAAGATTACTGTATTATTTTCAGATGTTTC									

	161210	161220	161230	161240	161250	161260	161270	161280	161290	161300
BAC-51									
BAC-51-15	GGGGGTATATTCTATTTAAGTTGAGTTTATAAGTGAGACATTGATTATAAACCTTTTCTCCTTCGTATTTATACGTACTAAGCTCTACATGACTACATT									
	161310	161320	161330	161340	161350	161360	161370	161380	161390	161400
BAC-51									
BAC-51-15	TTCATAGTTTGAAATATTTTCACACAGAATTCAAAAATAAACTTGTATCACAGTGTGGTGGGCGTTTCACAGTGTGGTAATCTCATACAATATAATGTGT									
	161410	161420	161430	161440	161450	161460	161470	161480	161490	161500
BAC-51									
BAC-51-15	CACAATGTAACGTAGTGGGCGTTTCACAGTGTGTTTGGTGTTCACAGTGTGGTGGGCGTTTCACAGTGTGGTAATCTCATACAATATAATGTGT									
	161510	161520	161530	161540	161550	161560	161570	161580	161590	161600
BAC-51									
BAC-51-15	TCACATTGTGGTTGCTTCGTTTACAGTGTGGTTCACAGTGTGGATATCTACACTGAAACGACGCGACCACAATGTGAAAATACCCCAATGTAACATTGT									
	161610	161620	161630	161640	161650	161660	161670	161680	161690	161700
BAC-51									
BAC-51-15	GGGCGTTTCACAGTGTGTTTCTGTTTCAAAGTGTGCTGGGCATTTCACTGTGTGAGTGTTTACAATGTTAAATTGGGGCGTTTCACATTGTGGTCGCTT									
	161710	161720	161730	161740	161750	161760	161770	161780	161790	161800
BAC-51									
BAC-51-15	CGTTTCACAGTTGGGTTTCACAGTATGAAACGACGCGACCACAATGTGAAAACGCCCAATTAAACATTGTGGGCGTTACACAGTGTGGTGGGCATTTTAC									
	161810	161820	161830	161840	161850	161860	161870	161880	161890	161900
BAC-51									
BAC-51-15	TGTGTGAGTGTTTACAATGTTAAATTGGGGCGTTTTCACAGTGTGATTACACTGTGTTTACACTGTGAATGAGTGTTGTGGTCTGGTCTGGTCGCGAC									

Alignment 2. BAC-52 and BAC-52-2b

	9910	9920	9930	9940	9950	9960	9970	9980	9990	10000
BAC52									
BAC52-2b	CATACTGAAT ACTAAAAATT GTAAAAATAA -GATGTAGCT GAACCTCATAC GAAGGTAATG TGCAATAATA AAGCCCCGAA TATTTTCCTG ACGCAGATGA									
	10010	10020	10030	10040	10050	10060	10070	10080	10090	10100
BAC52									
BAC52-2b	ATTGATCATT GTTATTGAAT CAATGAGGAA GACGAAATGA AGGAAAAAGG AATACAATGT ATATAGAGAT AGAGAGAGAG AGAGAGAGAG --ATAGAGAG									

	10110	10120	10130	10140	10150	10160	10170	10180	10190	10200
BAC52
BAC52-2b	GGATATATAT	ATATATATAT	ATATATATAG	AGAGAGAGAG	AGAGAGAGAG	AGA--GAATG	AGATAGAGAG	AAATAAAAGA	GAGAGAGAGA	GAA-GAAAGA
	10210	10220	10230	10240	10250	10260	10270	10280	10290	10300
BAC52
BAC52-2b	GAGAGG-AGA	GGTGAGATAG	ATAGAAAGGG	TAGAGAGAGA	TGAAGAGACA	AAAGAAATAG	AGATATGTAG	AGATAGAGAG	AGCGAGAGGG	AGAGACAAAG
	10310	10320	10330	10340	10350	10360	10370	10380	10390	10400
BAC52
BAC52-2b	AGACAGAGAG	ACAAAGAGAA	TATGAGAGTG	AGATGGAGAG	AGATATGAGT	GTGAGAGAAA	GAGG-AGAGA	GAGAGAGAGA	GAGAGAGAGA	GAGAAGATAG
	10410	10420	10430	10440	10450	10460	10470	10480	10490	10500
BAC52
BAC52-2b	ATAAGATTAT	TTATATATAC	AATGCGTATA	AAAAAAAGTG	TGCCCAACTT	TAGTAACCTC	TACTTAAAAA	TTTATAACAT	ATAAACTGAT	ATCCTGTCTA
	10510	10520	10530	10540	10550	10560	10570	10580	10590	10600
BAC52
BAC52-2b	ATGATTTTAT	ATTCATAATT	GTACTGCCAT	GGATGATTGA	GCAGCAGGCT	TTTTATAAAG	TTTTTTTCAA	ATCCTTTTTG	AACCAAGTTT	GGGCCGAAGC
	10610	10620	10630	10640	10650	10660	10670	10680	10690	10700
BAC52
BAC52-2b	AATGGATGCG	GGTCAAAAGT	CATTATGTGT	GGGTTATCTC	ATTCCATAAA	CAATTGTTCT	CTTATGAATA	CTAACTATTA	GGCTGTTGAA	GATAAAAGGT
	10710	10720	10730	10740	10750	10760	10770	10780	10790	10800
BAC52
BAC52-2b	GTGAATATCA	ACTCAAGCTA	TAAAGGAAAT	TTTATCACAA	ACAAAATGTA	TGATATTCTC	TTTGTTATGA	TGAATAAAAT	CTTAAATTCA	TTCGTATCCC
	10810	10820	10830	10840	10850	10860	10870	10880	10890	10900
BAC52
BAC52-2b	TTGTTTAAGG	AAACCTTTCT	CAGTGATAAA	AAACCGATCG	GGACGTTATC	TTCTTTTATTA	AAGTCTCATC	GTAATAATAA	TATGAAAAAT	TTATATAGCG
	10910	10920	10930	10940	10950	10960	10970	10980	10990	11000
BAC52
BAC52-2b	CTTGTGACAA	AAGTTTCAAA	GCACTCGTGT	GTTTCGTTCT	GCATTTGGAT	GTAGTAACCT	TTGAGTTTTT	ATTCGCTATT	CAATATAAAC	ACCATAATGT
	11010	11020	11030	11040	11050	11060	11070	11080	11090	11100
BAC52
BAC52-2b	GCTCATTACG	CGTGTGAAAT	TGTGTAGAGG	ACAAGTGAGC	AGAGAGAAAT	TAGATATAAT	GAAACAAGAA	GTTAGTTGGC	TGTTCAAAAT	AACTAAAAGT

	11110	11120	11130	11140	11150	11160	11170	11180	11190	11200
BAC52
BAC52-2b	TTTCCCCGGG	GTCGTAACAA	GGTTCTGCGA	AACAGGGGCG	GATCTAGCCG	GCGGCGAGGG	TGGGGGGGGG	GGGG----CA	ATTTAAGAAA	ATAGTTAGCG

	11210	11220	11230	11240	11250	11260	11270	11280	11290	11300
BAC52
BAC52-2b	CCGAATTAGC	CGGCGAAAAA	GCAATAGGGG	GGG--TTAAT	GAGCAATAAA	TTGTTATTTC	TTTTTTGCTC	AACGGGGGGG	GGGGTAGTGC	ATGCGCGGAT

Deletion 1, 5' end

	11310	11320	11330	11340	11350	11360	11370	11380	11390	11400
BAC52
BAC52-2b	CCAGGGGAGG	CCCCCGCCCC	CCCAAAAAAG	TTTTTAATTT	TTTGTTTTTT	TAAATGGAGA	CAGATAAAAA	TTTAGTGCTC	ACTACCACCC	CCCCCCCCCC

	11410	11420	11430	11440	11450	11460	11470	11480	11490	11500
BAC52
BAC52-2b	CCCCCTACTG	AGCAAAATTT	ATCGGCCGGC	ACGATTTTCG	AAT TTCACCA	CGCTAAATTA	AAAATTGATT	TCAAATTTGG	GTCTCCCCTA	AGGAATCTGT

	11510	11520	11530	11540	11550	11560	11570	11580	11590	11600
BAC52
BAC52-2b	GACCCGCGCC	AGTAGTAAGC	ACTATTTTTA	TTTACTGAAC	CCCCCCCCTT	GAGCAAAAAC	AAAAAGAAGG	TAAGATTGTG	GATACTATCT	TTGTTTAAAT

	119410	119420	119430	119440	119450	119460	119470	119480	119490	119500
BAC52
BAC52-2b	TGCCTGAAAA	AAAAATCACA	GTAGAAATAT	CCCCTGAAAA	GGATTGATCA	TCGTTTTTCGA	AATATTACCG	GAGGTAGCTG	GAAATTTGTT	GATAAATATT

	119510	119520	119530	119540	119550	119560	119570	119580	119590	119600
BAC52
BAC52-2b	CCATGCCACA	TGTGTAGTTT	AATATTAGCC	TACCCAATTT	CGTTTATGTG	TGCACTAAAA	AGTCATTTGT	CAGCATTTTC	AGACATTCTG	ATTAAATTC

	119610	119620	119630	119640	119650	119660	119670	119680	119690	119700
BAC52
BAC52-2b	TTGTAATTTT	GATGAATTCA	AACCTTACAC	TTCTGTACTA	AACAGGCGCG	TACGCAGGAT	TTTTTCAAGT	GGGGGGGGGG	GGGGGTTTAA	CATTTTTTAA

Deletion 1, 3' end

	119710	119720	119730	119740	119750	119760	119770	119780	119790	119800
BAC52
BAC52-2b	TCGGGCCGAA	AATTTTCGCAT	CGACTCAGCC	AGCCGCTGAA	TAAGCGGGGG	GGGGGGGGGG	GG--AAGAGG	ACACTTTTTC	TTTTCTTTT	TTGGTCTCGA

	119810	119820	119830	119840	119850	119860	119870	119880	119890	119900
BAC52
BAC52-2b	AATTTGAAAA	TTTGACATTT	TGCTCTGTTG	GGGGGGGGGG	GG-TAAGGTC	GCCTTTTTTTA	GGTCAGCCAT	GGGAATAGTT	TTTTT-ATTA	TTATTTTTTTT
	119910	119920	119930	119940	119950	119960	119970	119980	119990	120000
BAC52
BAC52-2b	TTT-ATTATT	CAAAAAACA	AAAAACA AAA	CAAAACAAAG	GGGGGGGGGG	TTGTTTTTTTT	AGGGGGGTTT	ATACACATAA	AAATACCAA	GGGGGGGGG-
	120010	120020	120030	120040	120050	120060	120070	120080	120090	120100
BAC52
BAC52-2b	TTAACCCCTA	AACACCCCC	CCCCCCC--T	GCGTACGCGC	CTGGTACTAC	ATACATATTA	AACCTTTCTA	TGTAAATTTT	AGCCCATTTC	TTTCAAAACG
	120110	120120	120130	120140	120150	120160	120170	120180	120190	120200
BAC52
BAC52-2b	TAATCACATG	ACCAGTACTA	GATTTATTTT	GAAACATCCG	TCTTGATGTT	TATAATGATA	GATGACGTTA	TACGACCCTG	TCTTATGGGG	TACCTTGTC
	120210	120220	120230	120240	120250	120260	120270	120280	120290	120300
BAC52
BAC52-2b	TTCGTGCCAT	TTTGGGGGGA	GGGTGGAGAC	CCGCAGACTT	CCCCCTCTAG	AACCACTACT	GAATACCACT	GAACGCAACT	GAAGAATATG	TTATCGTCAT
	120310	120320	120330	120340	120350	120360	120370	120380	120390	120400
BAC52
BAC52-2b	AGAGGTCGGT	TATACATTTT	TTTCCATGCA	GGTTTTCATT	CACTGTGAGG	TGCTCATTTG	TAATGATAAT	GACCCATCGT	CTCGGTGTTC	CCAGGGATGT
	120410	120420	120430	120440	120450	120460	120470	120480	120490	120500
BAC52
BAC52-2b	AAGTCTAGAT	TCAGACGTGG	TAGCCGTCAC	ACCCGTGGAG	CAAGCTCCAC	TCCTCATCTA	ATCTCTAATG	GACCTCTTTC	AACTGCCAC	ACTATGCATG
	120510	120520	120530	120540	120550	120560	120570	120580	120590	120600
BAC52
BAC52-2b	CCGCAGAAGC	ACACAACCT	GGTAATTACC	TTTTTAAAGA	ACACAACACT	TAACCTTAAA	GGTGTCGTGT	TTGTCTAGTG	AAAACGTCGT	TGGACTGTGG

Reference

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